

# Universal patterns in bone tissues: microstructure, composition, and mechanics

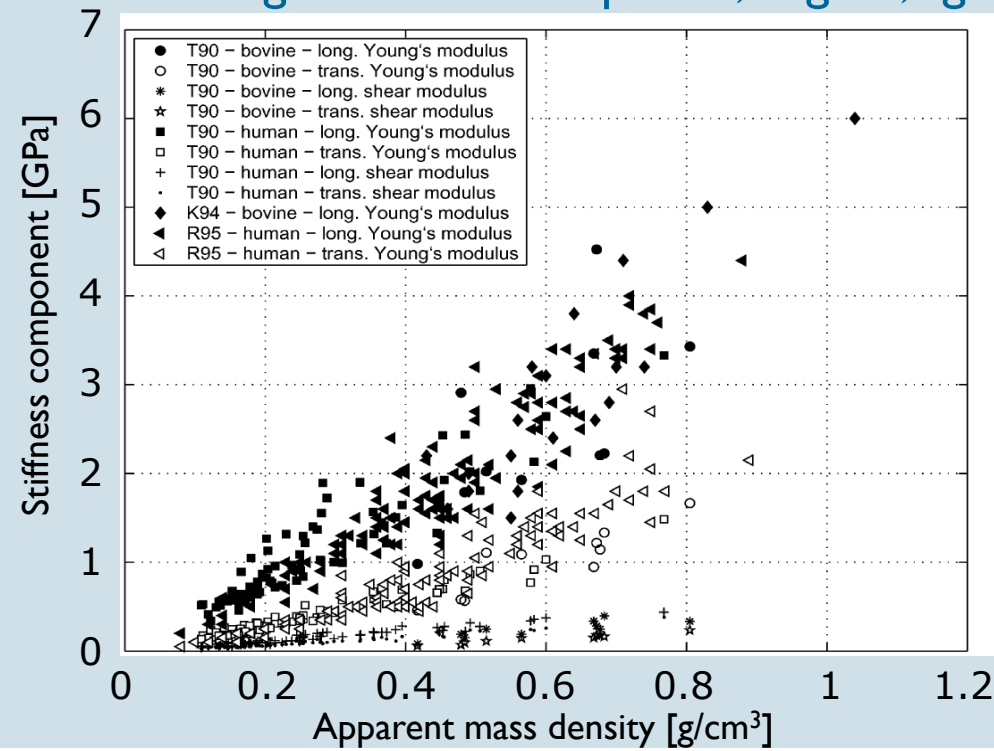
C. Morin<sup>1</sup>, P. Henits<sup>1</sup>, R. Blanchard<sup>1</sup>, Ch. Hellmich<sup>1</sup>

<sup>1</sup>Vienna University of Technology, Institute for Mechanics of Materials and Structures

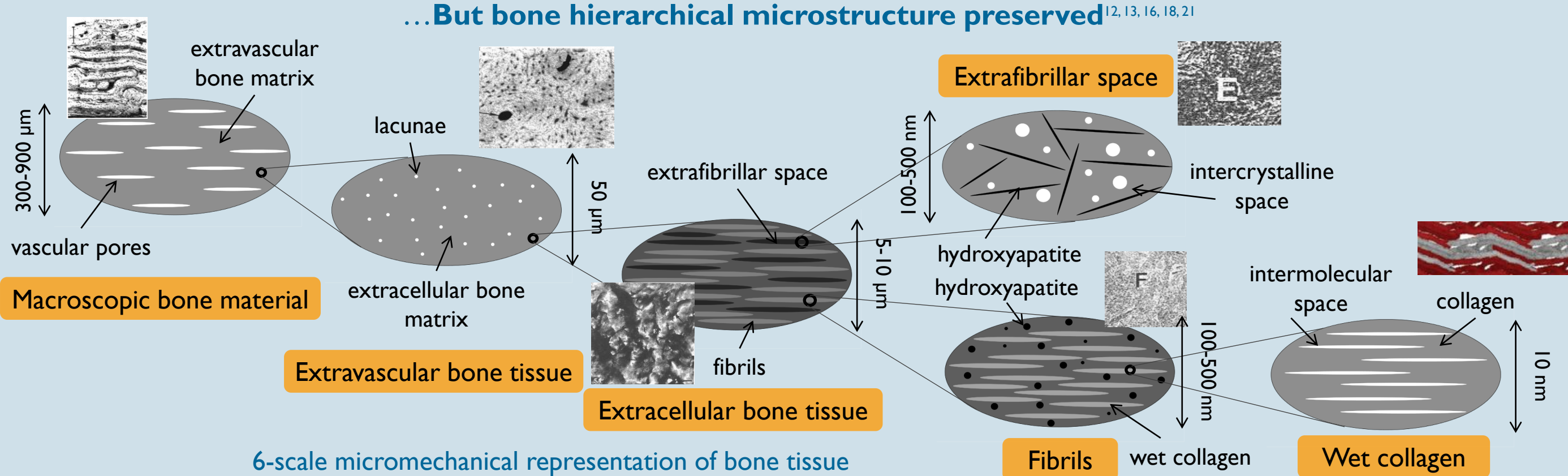
Supported by the European Commission and the European Research Council, ERC-2010-StG, 257023, MICROBONE.

## Motivation

Great diversity and variability of bone composition and mechanical behavior among the different species, organs, ages<sup>16</sup>, ...

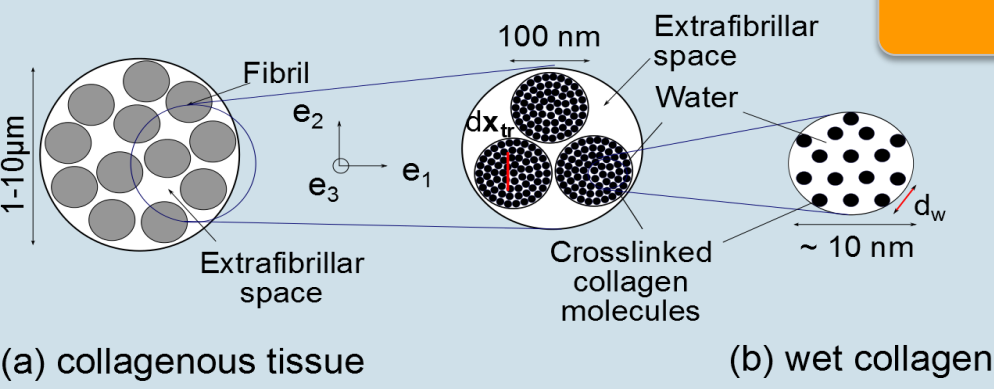


## Existence of universal patterns in bone tissues ?



## Structure of collagenous tissues

### Proportional swelling of fibrillar and extrafibrillar spaces<sup>39</sup>



Upon hydration, the extrafibrillar space growth evolves proportionally to the swelling of the fibrils.

$$V_{ef} = \beta (V_{fib} - V_{dry})$$

As soon as the fibrils are fully hydrated, only the extrafibrillar volume continues to grow.

The swelling rule is checked through...

mass-volume relation

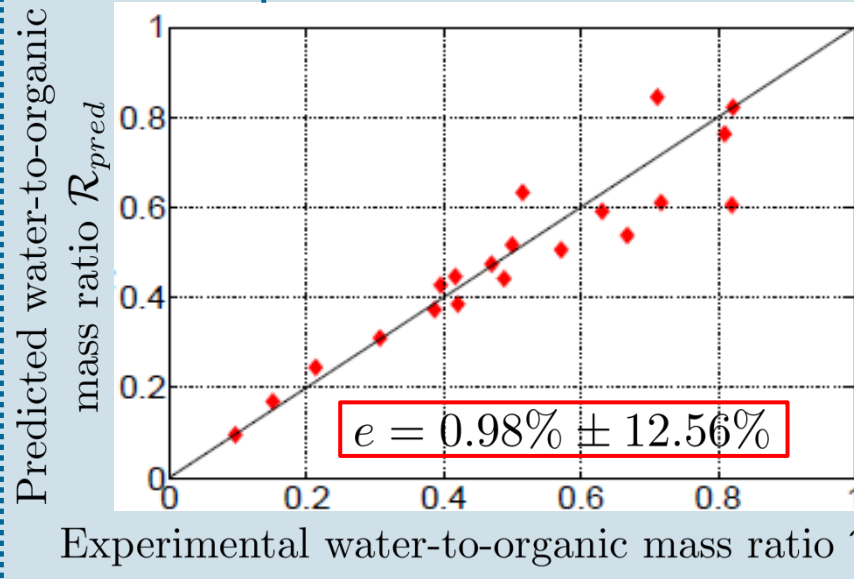
$$\frac{V_{tissue}}{V_{dry}} = 0.88 \frac{\mathcal{R} \rho_{col} + \rho_{H_2O}}{\rho_{H_2O}}$$

X-ray diffraction & continuum mechanics

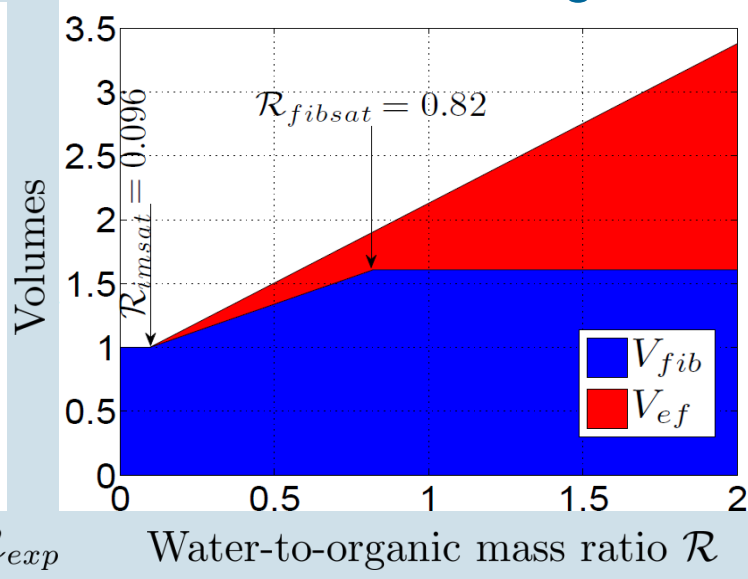
$$\frac{V_{fib}}{V_{dry}} = \left( \frac{d_w}{d_{dry}} \right)^2$$

$$\frac{V_{tissue}}{V_{dry}} = \beta \left( \frac{V_{fib}}{V_{dry}} - 1 \right) + \frac{V_{fib}}{V_{dry}}$$

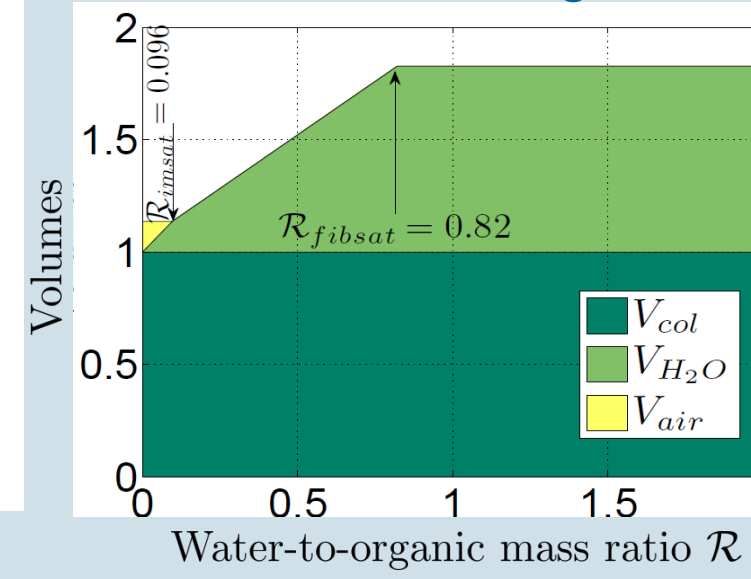
Experimental validation<sup>22,38,46</sup>



Tissue swelling

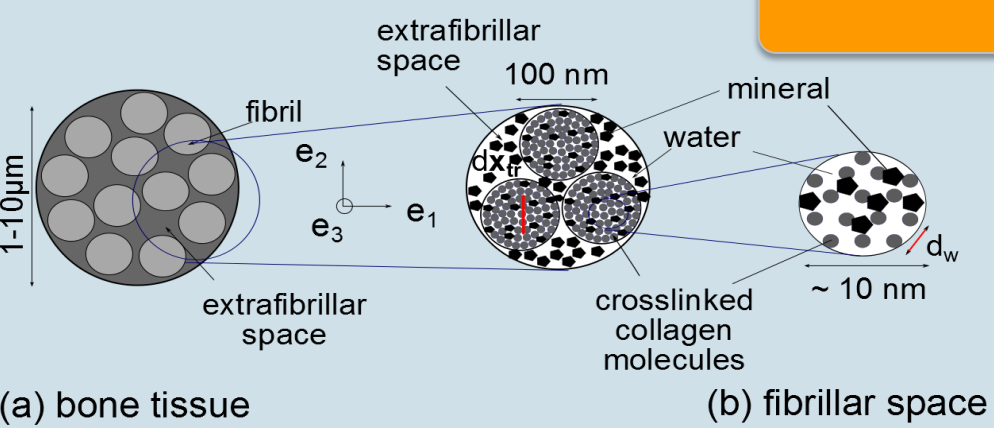


Fibrillar swelling



## Mineralization of collagenous tissues

### Mineralization occurs in closed thermodynamic systems<sup>40</sup>



Mineralizing bone tissue acts as a closed thermodynamic system, where crystals precipitate from an ionic solution:

$$\dot{m}_{fi}^{fib} = -\dot{m}_{HA}^{fib} \text{ and } \dot{m}_{fi}^{ef} = -\dot{m}_{HA}^{ef}$$

while the fibrillar and extrafibrillar masses are preserved:

$$m_{fib}^0 = m_{fib}^t = m_{fib}^\infty \text{ and } m_{ef}^0 = m_{ef}^t = m_{ef}^\infty$$

Translation of mass conservation to mass density-diffraction spacing relation by means of:

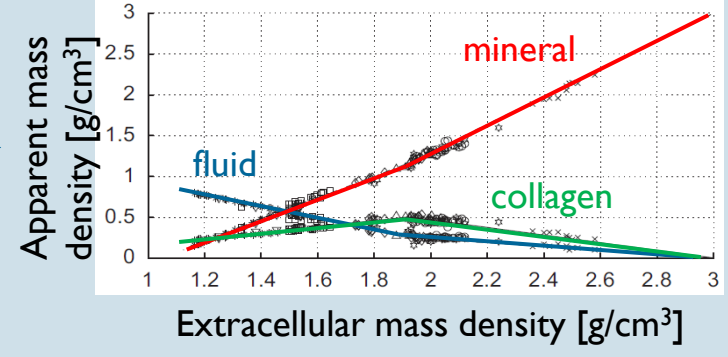
1. Bilinear laws for the composition of mineralized tissues<sup>1,3,14,15,26,27,31,34,48</sup>

2. Uniform extracollagenous mineral concentration<sup>19</sup>

3. Continuum deformation theory<sup>39</sup>

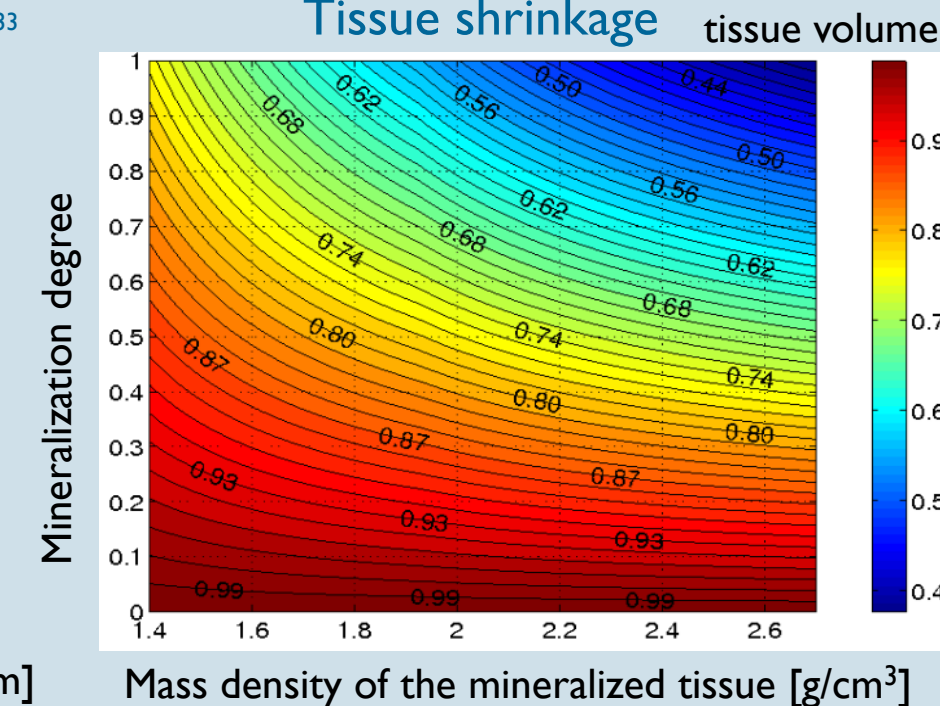
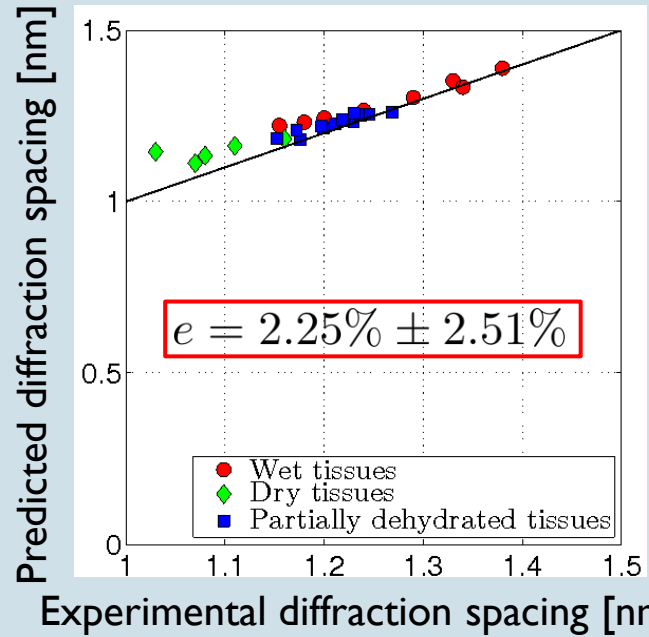
$$\frac{V_{fib}^\infty}{V_{fib}^0} = \left( \frac{d_w^\infty}{d_w^0} \right)^2$$

$$d_w^\infty = d_w^0 \sqrt{\frac{1 - f_{ef}^0}{1 - f_{ef}^\infty} \left[ 1 + \left( \frac{\rho_{HA}}{\rho_{fl}} - 1 \right) \frac{f_{HA}^\infty}{f_{HA}^0} \right]}$$



Check through the prediction of the diffraction spacing:

Experimental validation<sup>20,30-33</sup>

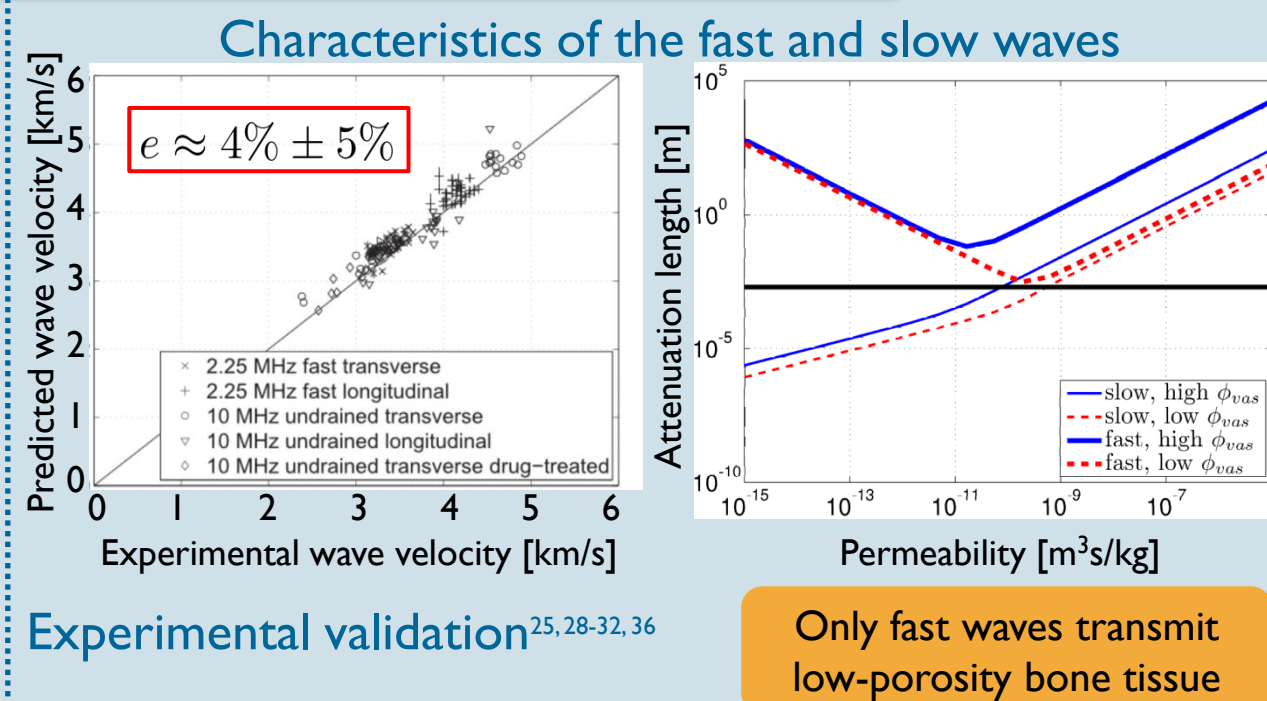
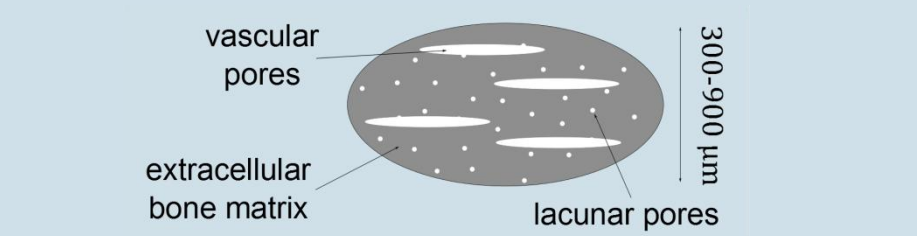


## Multiscale mechanical behavior of bone

### Wave propagation in bone tissues<sup>17,20,41</sup>

The characteristics of the longitudinal waves traveling in bone tissues are predicted by means of a **poro-micro-mechanical approach**<sup>17,20,48</sup>.

The micromechanical representation of the macroscopic bone tissue comprises two pore spaces (the *vascular pores*, allowing fluid flow and undrained *lacunar pores*), embedded in the *extracellular bone matrix*.



Experimental validation<sup>25,28-32,36</sup>

Only fast waves transmit low-porosity bone tissue

### Strength of bone tissues<sup>11,42</sup>

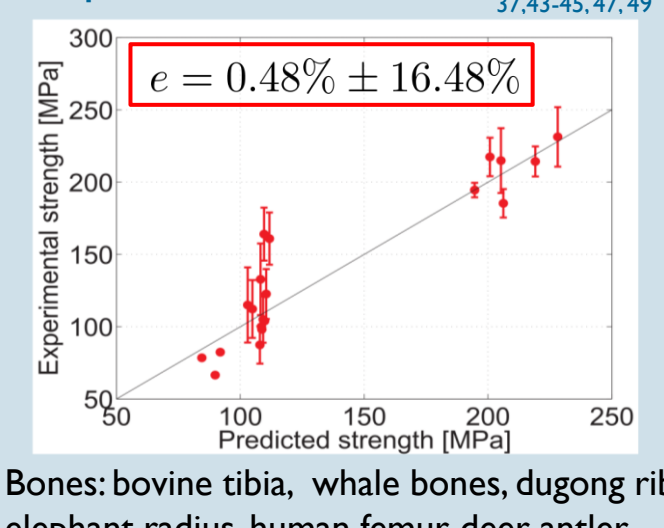
Bone failure can be explained by *mutual ductile sliding of mineral crystals along layered-water films*, modeled by an elastic-perfectly plastic law for hydrated mineral crystals

$$\mathcal{F}(\sigma) = k \max_i \sigma_{\theta,\phi}^i - \min_i \sigma_{\theta,\phi}^i - \sigma_{HA}^{ult} \leq 0$$

followed by *brittle failure of collagen crosslinks*

$$\mathcal{F}_{col}(\sigma_{col}) = \left| \max_i \sigma_{col,i} \right| - \sigma_{col}^{ult} \leq 0$$

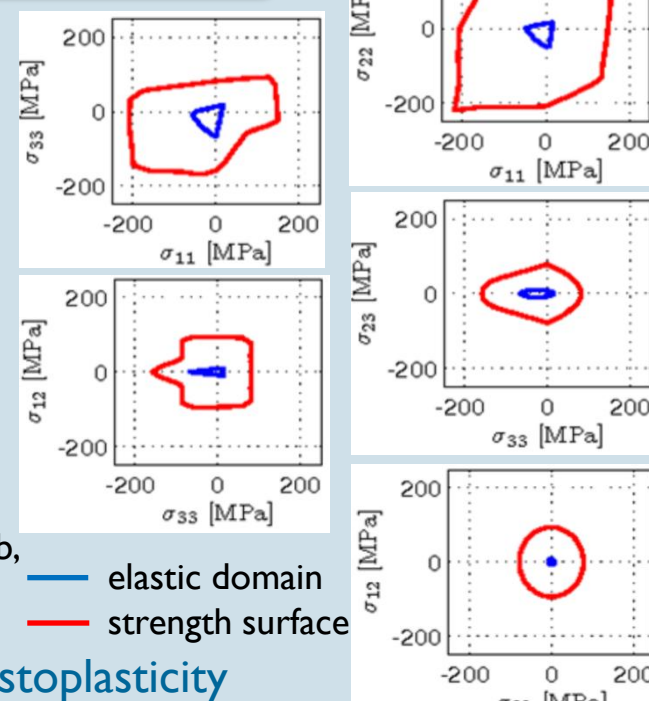
Experimental validation<sup>4-10,23-24,35,37,43-45,47,49</sup>



Bones: bovine tibia, whale bones, dugong rib, elephant radius, human femur, deer antler

— elastic domain — strength surface

Multiscale, multisurface elastoplasticity



## Conclusion

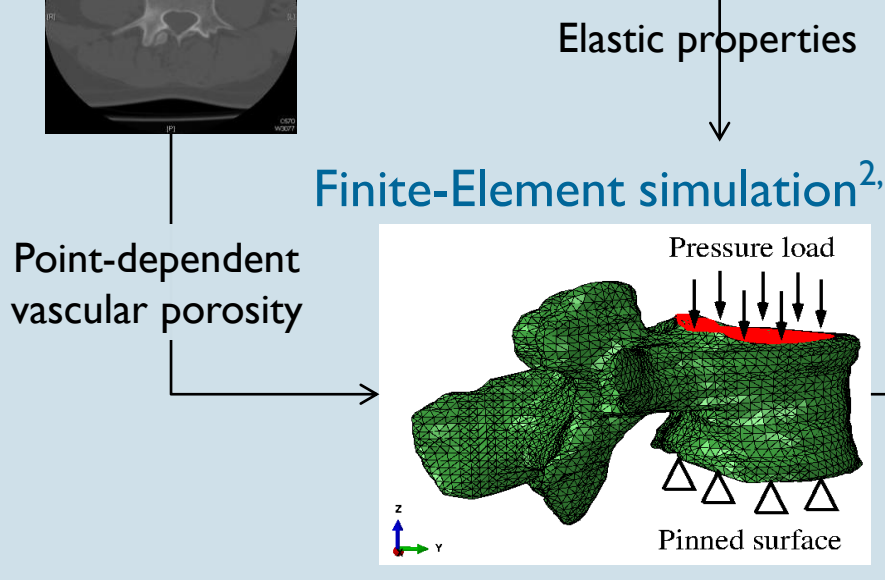
### Failure-risk analysis of a vertebra

CT images of a vertebra

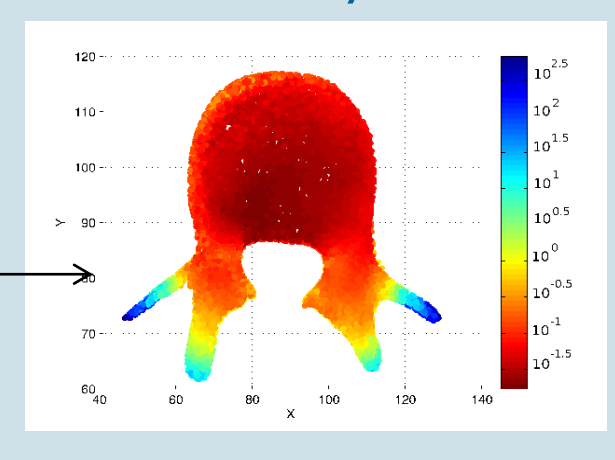
Micromechanical model

Elastic properties

Finite-Element simulation<sup>2,9</sup>



Post-treatment: distance to the yield surface



### Nomenclature

|                |                                                |            |                                         |
|----------------|------------------------------------------------|------------|-----------------------------------------|
| $d$            | equatorial diffraction spacing                 | $air$      | of air                                  |
| $f$            | volume fraction                                | $col$      | of collagen                             |
| $\mathcal{F}$  | yield function                                 | $dry$      | in dried tissues                        |
| $k$            | parameter of the Mohr-Coulomb criterion        | $ef$       | of extrafibrillar space                 |
| $m$            | mass                                           | $fib$      | of fibrillar space                      |
| $\mathcal{R}$  | water-to-organic mass ratio                    | $fibsat$   | at fibrillar saturation point           |
| $V$            | volume                                         | $fl, H_2O$ | of fluid                                |
| $\beta$        | proportionality constant                       | $HA$       | of hydroxyapatite                       |
| $\rho$         | mass density                                   | $imsat$    | at intermolecular pore saturation point |
| $\sigma$       | stress field                                   |            |                                         |
| $\sigma^i$     | i-th principal stress                          |            |                                         |
| $\sigma^{ult}$ | ultimate stress                                |            |                                         |
| $Superscripts$ |                                                |            |                                         |
| $ef$           | in the extrafibrillar space                    |            |                                         |
| $fib$          | in the fibrillar space                         |            |                                         |
| $0$            | at the end of the mineralization process       |            |                                         |
| $\infty$       | before the start of the mineralization process |            |                                         |

### Nomenclature and references

Indices  
of air  
of collagen  
in dried tissues  
of extrafibrillar space  
of fibrillar space  
at fibrillar saturation point  
of fluid  
of hydroxyapatite  
at intermolecular pore saturation point  
of collagenous tissue  
in wet tissues  
orientation angles of the hydroxyapatite needle in a spherical coordinate system

1. Blitz & Pellegrino, J. Bone Jt. Surg. 51-A (3), 456-466
2. Blanchard et al., J. Biomech. 45, 5478, 2012
3. Burns, Biochem. J. 23 (5), 860-867, 1929
4. Burstein et al., J. Biomech. 5, 35-44, 1972
5. Burstein et al., J. Bone Jt. Surg. 57-A, 956-961, 1975
6. Cezayirlioglu et al., J. Biomech. 18 (1), 61-69, 1985
7. Currey, J. Anatomy 93, 87-95, 1959
8. Currey, J. Biomech. 8, 81-86, 1975 & 23, 837-844, 1990 & 37, 549-556, 2004
9. Dejaco et al., J. Biomech. 45 (6), 1068-1075, 2012
10. Dickenson et al., J. Bone Jt. Surg. 63-B (2), 233-238, 1981
11. Fritsch et al., J. Theor. Biol. 260, 230-252, 2009
12. Fritsch & Hellmich, J. Theor. Biol. 244, 597-620, 2007
13. Fritsch et al., J. Biomed. Mater. Res. A 88, 149-161, 2009
14. Gong, et al., Anat. Rec. 149, 325-332, 1964
15. Hammett, J. Eng. Mech. A 23, 783-810, 2004
16. Hellmich et al., Eur. J. Mech. A 23, 783-810, 2004
17. Hellmich et al., J. Eng. Mech. 135 (5), 382-394, 2009
18. Hellmich et al., Biomech. Model. Mechanobiol. 2(4), 219-238, 2004
19. Hellmich & Ulm, Biomech. Model. Mechanobiol. 2 (1), 21-36, 2003
20. Hellmich & Ulm, J. Eng. Mech. 131 (9), 918-927, 2005
21. Hellmich & Ulm, J. Biomech. 35, 1199-1212, 2002
22. Katz & Li, J. Med. Biol. 80 (1), 1-15, 1973
23. Kotha & Guzelsu, J. Theor. Biol. 219, 269-279, 2002
24. Lee et al., Bone 21, 119-125, 1997
25. Lees, J. Biomech. 15 (11), 867-874, 1982
26. Lees, Biophys. J. 85, 204-207, 2003
27. Lees et al., J. Acoust. Soc. Am. 66 (3), 641-646, 1979
28. Lees et al., J. Bone Miner. Res. 9 (9), 1377-1389, 1994
29. Lees et al., J. Acoust. Soc. Am. 74 (1), 28-33, 1983
30. Lees et al., Int. J. Macromol. 6 (6), 321-326, 1984
31. Lees et al., J. Acoust. Soc. Am. 99 (4), 2421-2427, 1995
32. Lees et al., Calc. Tissue Int. 29, 107-117, 1979
33. Lees & Mook, Calc. Tissue Int. 39, 291-292, 1986
34. Lees & Page, Connect. Tissue Res. 28 (4), 263-287, 1992
35. McCalden et al., J. Bone Jt. Surgery 75-A, 1193-1205, 1993
36. McCarthy et al., J. Biomech. 22 (11), 1139-1143, 1990
37. Martin & Ishida, J. Biomech. 22, 419-426, 1989
38. Meek et al., Biophys. J. 60 (2), 467-474, 1991
39. Morin & Hellmich, J. Theor. Biol. 317, 384-393, 2013
40. Morin & Hellmich, submitted to Ultrasonics, 2013
41. Pichler & Hellmich, J. Eng. Mech. 136 (8), 1043-1053, 2010
42. Reilly & Burstein, J. Biomech. 22, 419-426, 1989
43. Reilly & Burstein, J. Biomech. 8, 393-405, 1975
44. Riggs et al., Anat. Embryol. 187, 239-248, 1993
45. Rougier & Bear, J. Am. Leather Chem. Ass. 48, 735-751, 1953
46. Sedlin & Hirsch, Acta Orthop. Scand. 37, 29-48, 1966
47. Vuong & Hellmich, J. Theor. Biol. 287, 115-130, 2011
48. Walsh & Guzelsu, Biomat. 15 (2), 137-145, 1994